Effectiveness of Xtreme Bio in Deactivating Porcine Circovirus 2 (PCV-2)

Dr. Jianqiang Zhang, PhD

Project Title: Evaluation of *in vitro* antiviral effect of the disinfectant XTREME BIO on PCV2

Project Objectives:

The objectives of this project was to evaluate the *in vitro* antiviral effect of XTREME BIO on porcine circovirus 2 [PCV-2].

Procedures to Achieve Objectives:

Materials

<u>Liquid compounds from Midwest Ag Solutions, LLC:</u> Xtreme Bio (1 gallon).

Virus: A PCV2d isolate (USA/NC24897/2016, P6) that grew efficiently in PK-15 cells.

Cells: Porcine kidney (PK-15) cells (ATCC CCL-33).

<u>Cell culture maintenance medium and inoculation medium</u>: MEM supplemented with 10% fetal bovine serum, 2 mM L-glutamine, 0.05 mg/ml gentamicin, 10 unit/ml penicillin, 10 μ g/ml streptomycin, and 0.25 μ g/ml amphotericin.

<u>D-glucosamine:</u> 300 mM D-glucosamine.

PCV2 FA conjugate: VMRD CJ-F-PCV2-10ML.

Culture plates, serological pipette, tips and other supplies.

Methods

1. Test cytotoxicity of the product Xtreme Bio in PK-15 cells.

The recommended dilution of Xtreme Bio is 0.5 oz/gallon (1:256) on clean surface and 2 oz/gallon (1:64) in rough surface. (1 gallon = 128 oz).

In this study, the product Xtreme Bio was serially diluted in autoclaved distilled water: 1:16, 1:32, 1:64, 1:128, 1:256, and 1:512.

- 1) Prepare PK-15 cells in 96-well plates one day before the cytotoxicity testing.
- 2) The serially diluted product was inoculated in triplicate onto sub-monolayer of PK-15 cells seeded the previous day in a 96-well plate (100 μ l of product per well). Controls included autoclaved water for inoculation. After 1 h incubation at 37°C, the inoculum was removed and 100 μ l of fresh medium per well was added.
- 3) After incubation at 37°C for 24-48 h, the cells were scraped and harvested into 1 ml culture medium. Determine the cell density and percentage of live and dead cells using trypan blue staining and hemocytometer or ViCell XR (Beckman Coulter).
- 4) The 50% cytotoxic concentration (CC_{50}) were defined as the product dilution that caused reduction of 50% viability compared to the water-inoculated cells.

2. Treatment of PCV2 with the product Xtreme Bio (Table 1)

- 1) The product Xtreme Bio was serially diluted in autoclaved distilled water: 1:16, 1:32, 1:64, 1:128, 1:256, and 1:512.
- 2) The serially diluted Xtreme Bio product (300 μ l) was mixed with the equal volume of PCV2d isolate (300 μ l, 10^4-5 TCID50/ml) in 5 ml sterile snap-cap tubes (triplicate for each group). Control groups include water + virus, and water + medium. In total, 8 groups with 3 replicates for each group (treatments #1-24).
- 3) After incubation at room temperature (20-25°C) for 30 min, each sample was titrated for PCV2 in PK-15 cells. Briefly, serial 10-fold dilutions of each sample (10^0, 10^-1, 10^-2 and 10^-3) were prepared from each mixture and then inoculated into the sub-monolayers of PK-15 cells grown in 96-well plates (100 μ l per well, 2 wells per dilution). Incubate at 37°C for 1 h. Discard the supernatant (inoculum). Add fresh medium (100 μ l per well).
- 4) The plates were incubated at 37°C in 5% CO2 incubator for 24 h. Check if any cytotoxicity occurs. Add 50 μ l of 300 mM D-glucosamine to each well of 96-well plates and incubate (with plates covered in dark) at room temperature for approximately 15-20 min. Decant D-glucosamine. Wash plate wells with 100 μ l MEM/10% FBS and decant the medium. Add 100 μ l MEM/10% FBS to each well.
- 5) Incubate plates for additional 24h. Check cells condition. Transfer the supernatants to new 96-well plates and save the supernatants. Rinse the cell plates once with PBS and fix the plates with 80% cold acetone for at least 10 min. Allow the fixed plates to dry. Add 50 μl of PCV2 FA conjugate from VMRD. Incubate at 37°C for about 1 h. Decant conjugate and wash plates with PBS for 3 times, about 5 min per time. Read plates under fluorescence microscope.
- 6) For each treatment (#1-24), pick one appropriate dilution (10^-1) of the supernatants for quantitative PCV2 PCR (pool duplicate wells of supernatant at that dilution).
- 7) Data analysis.

Table 1. Experimental design of evaluating antiviral effect of Xtreme Bio on PCV2

The mixture of PCV2 and Xtreme BioShield were serially diluted $(10^0, 10^{-1}, 10^{-2} \text{ and } 10^{-3})$ for titration in PK-15 cells. Triplicate for each group. The supernatants from the 10^{-1} dilutions collected from 48 h post-inoculation into PK-15 cells were tested by quantitative PCV2 PCR.

Treatment	PCV2 PCR Ct value				PCV2 genomci copies/ml				Percentage of genomic copies
	Replicate 1	Replicate 2	Replicate 3	Mean	Replicate 1	Replicate 2	Replicate 3	Mean	reduction
PCV2 + water	20.2	19.5	19.8	19.8	1.16E+08	1.81E+08	1.48E+08	1.48E+08	0.00%
PCV2 + Xtreme BioShield 1:16	27.0	26.2	26.0	26.4	1.33E+06	2.32E+06	2.60E+06	2.08E+06	98.60%
PCV2 + Xtreme BioShiel 1:32	24.5	24.4	26.2	25.0	7.01E+06	7.35E+06	2.30E+06	5.55E+06	96.26%
PCV2 + Xtreme BioShiel 1:64	22.8	23.3	22.5	22.9	2.15E+07	1.48E+07	2.63E+07	2.09E+07	85.93%
PCV2 + Xtreme BioShiel 1:128	22.9	22.6	21.9	22.5	1.97E+07	2.38E+07	3.80E+07	2.71E+07	81.71%
PCV2 + Xtreme BioShiel 1:256	21.0	20.1	21.8	21.0	6.61E+07	1.20E+08	3.91E+07	7.51E+07	49.39%
PCV2 + Xtreme BioShiel 1:512	21.2	20.6	20.7	20.8	5.90E+07	9.11E+07	8.49E+07	7.83E+07	47.19%
Medium + water	34.1	32.5	>37						

Percentage of genomic copies reduction = (untreated PCV2 genomic copies - treated PCV2 genomic copies) / (untreated PCV2 genomic copies) X 100%

Results:

1. Cytotoxicity of the product Xtreme Bio in PK-15 cells.

The product Xtreme Bio at all of the tested dilutions 1:16, 1:32, 1:64, 1:128, 1:256, and 1:512 led to detachment of PK-15 cells. It appears that Xtreme Bio at these dilutions was cytotoxic to PK-15 cells.

2. Evaluation of in vitro antiviral effect of the product Xtreme Bio on PCV2

The product Xtreme Bio was serially diluted in autoclaved distilled water: 1:16, 1:32, 1:64, 1:128, 1:256, and 1:512. Then the serially diluted Xtreme Bio product (300 μ l) was mixed with the equal volume of PCV2d isolate (300 μ l) and incubated at room temperature (20-25°C) for 30 min followed by titration in PK-15 cells.

The records of cell conditions and PCV2 immunofluorescence staining results are shown in the attached Excel spreadsheet 'CPE and FA'. For the negative control (virus-negative culture medium treated with water), all of the wells were PCV2 FA staining negative. For the positive control (virus treated with water), all of the wells inoculated with 10^{0} , 10^{-1} , 10^{-2} , and 10^{-3} dilutions of the virus + water mixture had positive FA staining and the virus titer in this group was $10^{4.5}$ TCID50/ml. When PCV2 was treated with Xtreme Bio at 1:16, 1:32, and 1:64 dilutions, the 10^{0} and 10^{-1} dilutions of the mixture were PCV2 FA staining negative; thus no virus titer can be calculated. When PCV2 was treated with Xtreme Bio at 1:128, 1:256, and 1:512 dilutions, the 10^{0} dilutions of the mixture led to cell detachment, the 10^{-1} dilutions of the mixture were FA staining positive and the 10^{-2} and 10^{-3} dilutions of the mixture were PCV2 FA staining negative; this gave rise to virus titer of $10^{2.5}$ TCID50/ml. Thus, compared to the untreated virus, treatment with the Xtreme Bio at dilutions of 1:128, 1:256 and 1:512 resulted in 2 log10 reduction of the infectious virus titer.

For each treatment, the supernatants from the 10⁻¹ dilutions (triplicate for each treatment) of the mixture after 48 h inoculation in PK-15 cells were tested by a quantitative PCV2 PCR. The results are summarized in the Excel spreadsheet 'PCR'. Compared to the untreated PCV2 group, the treatment with Xtreme Bio at 1:16, 1:32, 1:64, 1:128, 1:256 and 1:512 dilutions resulted in 98.60%, 96.26%, 85.93%, 81.71%, 49.39%, and 47.19% reduction of genomic copies, respectively.

Conclusion:

Under the conditions of this study, compared to the untreated PCV2, treatment with the Xtreme Bio at dilutions of 1:128, 1:256 and 1:512 resulted in 2 log10 reduction of the infectious virus titer. Compared to the untreated PCV2 group, the treatment with Xtreme Bio at 1:16, 1:32, 1:64, 1:128, 1:256 and 1:512 dilutions resulted in 98.60%, 96.26%, 85.93%, 81.71%, 49.39%, and 47.19% reduction of genomic copies, respectively.